# Niphatesines E-H, New Pyridine Alkaloids from the Okinawan Marine Sponge Niphates sp. 

Jun'ichi Kobayashi,*, a Chun-min Zeng, ${ }^{a}$ Masami Ishibashi, ${ }^{a}$ Hideyuki Shigemori, ${ }^{a}$<br>Takuma Sasaki ${ }^{\text {b }}$ and Yuzuru Mikami ${ }^{\text {c }}$<br>${ }^{a}$ Faculty of Pharmaceutical Sciences, Hokkaido University, Sapporo 060, Japan<br>${ }^{\text {b }}$ Cancer Research Institute, Kanazawa University, Kanazawa 920, Japan<br>${ }^{c}$ Research Center for Pathogenic Fungi and Microbial Toxicoses, Chiba University, Chiba 280, Japan

Four new mono-3-alkyl-substituted pyridine alkaloids, niphatesines $\mathrm{E}-\mathrm{H}$ (1-4), possessing oxime methyl ether or methoxyamine functionality have been isolated from the Okinawan marine sponge Niphates sp. and the structures elucidated by spectral and chemical means. These pyridine alkaloids exhibited cytotoxic and antimicrobial activities.

During our continuing studies on bioactive substances from Okinawan marine organisms, ${ }^{1}$ we previously described the isolation and structure elucidation of niphatesines A-D, monosubstituted pyridine alkaloids possessing antileukaemic activity from an extract of the Okinawan sponge Niphates sp. ${ }^{2}$ Further investigation of the constituents of this sponge has now led to the isolation of four new pyridine alkaloids, niphatesines E-H (1-4), with cytotoxic and antimicrobial activities. Here we describe the isolation and structure elucidation of these new alkaloids 1-4.

The sponge Niphates sp. was collected off the Kerama Islands, Okinawa, and kept frozen until used. The methanol extract was partitioned between ethyl acetate and water. The ethyl acetatesoluble fraction was subjected to repeated silica gel flash column chromatography and elution with chloroform-methanol and hexane-ethyl acetate, followed by reversed-phase HPLC to give niphatesines E (1, $0.0005 \%$ yield, wet weight), F ( $\mathbf{2}, \mathbf{0 . 0 0 1 \% \text { ), G }}$ $(\mathbf{3}, 0.001 \%)$, and $\mathrm{H}(4,0.002 \%)$ together with the known ikimine A (5, 0.002\%). ${ }^{3}$

The molecular formula of niphatesine E (1) was determined to be $\mathrm{C}_{20} \mathrm{H}_{30} \mathrm{~N}_{2} \mathrm{O}$ by high-resolution fast-atom bombardment mass spectrometry (HR-FAB-MS) data [ $\mathrm{m} / \mathrm{z} 315.2430$ (M + $H)^{+}, \Delta-0.6 \mathrm{mmu}$ ]. The UV absorption maximum at 262 nm implied the presence of the same pyridine chromophore as that of niphatesines A-D. ${ }^{2}$ The ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals for the aromatic region $\left[\delta_{\mathrm{H}} 8.84(\mathrm{br} \mathrm{s}, 2-\mathrm{H}), 8.79(\mathrm{~d}, J 5.9 \mathrm{~Hz}, 6-\mathrm{H}), 8.29\right.$ (d, $J 8.1 \mathrm{~Hz}, 4-\mathrm{H}$ ) and 7.83 ( $\mathrm{dd}, J 8.1$ and $5.9 \mathrm{~Hz}, 5-\mathrm{H}$ ); $\delta_{\mathrm{C}} 144.4$ (d, C-4), 142.7 (s, C-3), 140.7 (d, C-2), 140.5 (d, C-6) and 125.9 (d, C-5)] also suggested the presence of a mono-3-alkyl-substituted pyridine ring on the basis of comparison with spectral data of niphatesines ${ }^{2}$ and theonelladins. ${ }^{4}$ The ${ }^{13} \mathrm{C}$ NMR signals at $\delta_{\mathrm{C}}$ 83.9 (s) and 76.0 (s; overlapped with the signal of $\mathrm{CDCl}_{3}$ ) were indicative of the presence of a disubstituted alkyne moiety. Six degrees out of the seven unsaturation numbers were accounted for by the pyridine ring and alkyne group. In the ${ }^{1} \mathrm{H}$ NMR spectrum of compound 1 a methoxy and an olefinic proton were observed as split signals in the ratio 1.4:1 ( $\delta_{\mathrm{H}} 3.81$ and 3.87 , each singlet; $\delta_{\mathrm{H}} 7.36$ and 6.66 , each triplet, $J 6.5 \mathrm{~Hz}$, respectively). The ${ }^{13} \mathrm{C}$ signals at $\delta_{\mathrm{C}} 152.1$ and $151.1(\mathrm{C}-20)$ and $\delta_{\mathrm{C}} 61.6$ and 6.1.2 ( OMe ) also appeared as split signals in ratios $\sim 1.4: 1$. These ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ signals were assignable to an oxime methyl ether ( $-\mathrm{CH}=\mathrm{NOMe}$ ) present at the terminus of the alkyl chain, existing as a mixture of $E$ and $Z$ isomers. The major isomer was assigned as $E$ on the basis of the ${ }^{1} \mathrm{H}$ chemical shifts ( $\delta_{\mathrm{H}} 7.36$ and $6.66,1.4: 1,20-\mathrm{H}$ ), since the proton attached to the oxime carbon of the $E$-isomer usually resonates at lower field than does that of the $Z$-isomer. ${ }^{5}$ The presence of the oxime methyl ether group was further confirmed by the intense fragment ion

peaks observed at $m / z 283\left(\mathrm{M}-\mathrm{OCH}_{3}\right)^{+}$and $256(\mathrm{M}-\mathrm{CH}=$ $\mathrm{NOMe}^{+}$in the electron-impact mass spectrum (EIMS) of compound 1 as well as the IR absorption band of compound 1 at $1680 \mathrm{~cm}^{-1}$. The remaining one unsaturation was thus characterized. The position of the alkyne group of compound 1 was deduced to be at C-9 and C-10 from the EIMS fragmentation pattern (Fig. 1), which was virtually the same pattern as that of niphatesine A. ${ }^{2}$ The structure of niphatesine E was, therefore, concluded to be 1 .

1


Fig. 1 The EIMS fragment ions ( $m / z$ of niphatesines E 1 and H 4
Niphatesine F (2) was shown to have the molecular formula $\mathrm{C}_{22} \mathrm{H}_{34} \mathrm{~N}_{2} \mathrm{O}$ by HR-FAB-MS data [ $\mathrm{m} / \mathrm{z} 343.2750(\mathrm{M}+\mathrm{H})^{+}$, $\Delta+0.1 \mathrm{mmu}]$. The spectral data of niphatesine F (2) resembled closely those of niphatesine $E$ (1) except for the molecular weight, suggesting that compound 2 contained two more methylene units ( $28 \mathrm{amu} ; \mathrm{CH}_{2} \times 2$ ) in the side-chain compared with compound 1. A methoxyimino group was also revealed to be present at the terminus of the side-chain by ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectroscopy of a mixture of $E$ and $Z$ isomers in the ratio 1.7:1 $\left(\delta_{\mathrm{H}} 3.81\right.$ and 3.86, each singlet, $\mathrm{OMe} ; \delta_{\mathrm{H}} 7.36$ and 6.66, each triplet, $J 6.2 \mathrm{~Hz}, 22-\mathrm{H} ; \delta_{\mathrm{C}} 61.5$ and $61.2, \mathrm{OMe} ; \delta_{\mathrm{C}} 151.9$ and 150.9, C-22, respectively), the major one again being the $E$ isomer. The position of the alkyne group was clearly revealed to be at C-15 and C-16 on the basis of a comparison of the EIMS fragmentation pattern with that of niphatesine $\mathrm{B}^{2}[\mathrm{~m} / \mathrm{z} 228$ (fission of $\mathrm{C}-17 / \mathrm{C}-18$ bond) and 176 (fission of $\mathrm{C}-13 / \mathrm{C}-15$ bond)]. Thus the structure of niphatesine F was assigned to be 2 .
Niphatesine G (3) was also a mono-3-alkyl-substituted pyridine alkaloid with an alkyl side-chain terminated by an oxime methyl ether group. The molecular formula of compound $\mathbf{3}$ was established to be $\mathrm{C}_{20} \mathrm{H}_{34} \mathrm{~N}_{2} \mathrm{O}$ by HR-FAB-MS data [3: m/z $\left.319.2743(\mathrm{M}+\mathrm{H})^{+}, \Delta-0.6 \mathrm{mmu}\right]$. The ${ }^{1} \mathrm{H}$ NMR spectrum of compound 3 was quite similar to that of ikimine A (5, 2.7:1 mixture of $E / Z$ isomers), which was also isolated from this sponge. Ikimine A (5) was previously isolated from an unidentified Micronesian sponge. ${ }^{3}$ The ${ }^{1} \mathrm{H}$ NMR spectrum of compound $\mathbf{3}$ revealed signals due to a secondary methyl group ( $\delta_{\mathrm{H}} 1.05,3 \mathrm{H}, \mathrm{d}, J 6.6 \mathrm{~Hz}$ ) as well as the proton attached to the oxime carbon, observed as a doublet ( $\delta_{\mathrm{H}} 7.20$ and $6.44, J 7.7 \mathrm{~Hz}$, $19-\mathrm{H} ; E: Z 3.2: 1$ ). The difference in the spectral properties of compounds 3 and 5 was found only in the molecular weight [HR-FAB-MS of 5: m/z $305.2585(\mathrm{M}+\mathrm{H})^{+}, \mathrm{C}_{19} \mathrm{H}_{33} \mathrm{~N}_{2} \mathrm{O}, \Delta$ $-0.8 \mathrm{mmu}]$. Compound 3 was, therefore, shown to possess one more $\mathrm{CH}_{2}$ unit ( 14 amu ) between the 3-position of the pyridine ring and the terminal methoxy imino group than has compound 5. The presence of the oxime methyl ether group in compound 5 was confirmed by the following chemical tests: on treatment with $3 \mathrm{~mol} \mathrm{dm}^{-3}$ hydrochloric acid in methanol under reflux for 20 h , compound 5 was converted into the corresponding aldehyde $\left[6, m / z 275\left(\mathrm{M}^{+}\right)\right.$and $246(\mathrm{M}-\mathrm{CHO})^{+}$]. The ${ }^{1} \mathrm{H}$

NMR spectrum of the aldehyde 6 no longer exhibited the signal due to a methoxy group, but instead the signal due to an aldehyde group was observed at $\delta_{\mathrm{H}} 9.61$ as a doublet $(J 1.9 \mathrm{~Hz})$.

The molecular formula of niphatesine $\mathrm{H}(4), \mathrm{C}_{20} \mathrm{H}_{32} \mathrm{~N}_{2} \mathrm{O}$, established by HR-FAB-MS $\left[m / z 317.2600(\mathrm{M}+\mathrm{H})^{+}, \Delta+0.7\right.$ mmu ], implied that niphatesine $\mathbf{H}$ (4) possesses two more hydrogen atoms than does niphatesine $\mathrm{E}(\mathbf{1})$. The UV absorption of compound 4 ( $\lambda_{\text {max }} 262 \mathrm{~nm}$ ) was similar to that of compound 1 while the IR spectrum of compound 4 did not show the absorption band at $1680 \mathrm{~cm}^{-1}$, thus suggesting the presence of a pyridine chromophore and the absence of the oxime group in compound 4 . The ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectra of compound 4 revealed signals due to a methoxy $\left[\delta_{\mathrm{H}} 3.93(3 \mathrm{H}, \mathrm{s}) ; \delta_{\mathrm{C}} 61.4(\mathrm{q})\right]$, a disubstituted alkyne $\left[\delta_{\mathrm{C}} 84.2\right.$ (s) and 76.0 ( s ; overlapped with the signal of $\left.\mathrm{CDCl}_{3}\right)$ ], and a heteroatom-bearing sp ${ }^{3}$ methylene [ $\left.\delta_{\mathrm{H}} 3.26(2 \mathrm{H}, \mathrm{br} \mathrm{s}) ; \delta_{\mathrm{C}} 49.4(\mathrm{t})\right]$ unit. These spectral features were parallel to those of niphatyne A (7), ${ }^{6}$ previously isolated from a Fijian sponge (Niphates sp.), and compound 4 was, therefore, inferred to be a 3 -alkylpyridine alkaloid with a methoxy amino group at the terminus of the alkyl chain. The position of the alkyne functionality of compound 4 was deduced to be at C-9 and C-10 on the basis of comparison of the EIMS data with those of niphatesine E (1) as shown in Fig. 1. Thus niphatesine H (4) was shown to be the $20, \mathrm{~N}$-dihydro form of niphatesine $E$ (1). The oxime bond of niphatesine $E$ (1) was veritably reduced by treatment with sodium cyanoborohydride to afford niphatesine H (4) almost quantitatively. By the same procedure niphatesine F (2) was converted into known niphatyne A (7), ${ }^{6}$ which was also isolated from this sponge, to verify the structure of compound 2.

Niphatesines E-H (1-4) and compound 5 exhibited cytotoxic activity against murine lymphoma L1210 cells and human epidermoid carcinoma KB cells in vitro. The inhibitory activity (\%) at $10 \mu \mathrm{~g} / \mathrm{cm}^{3}$ against L1210 and KB cells, respectively, were as follows: $\mathbf{1 :} 27.0 \%$ and $22.7 \%, \mathbf{2}: 41.2 \%$ and $19.4 \%, \mathbf{3}:>50 \%$ $\left(\mathrm{IC}_{50} 7.9 \mu \mathrm{~g} / \mathrm{cm}^{3}\right.$ ) and $16.8 \%$, 4: > 50\% ( $\mathrm{IC}_{50} 1.9 \mu \mathrm{~g} / \mathrm{cm}^{3}$ ) and $>50 \%\left(\mathrm{IC}_{50} 6.0 \mu \mathrm{~g} / \mathrm{cm}^{3}\right)$, and $5:>50 \%\left(\mathrm{IC}_{50} 5.4 \mu \mathrm{~g} / \mathrm{cm}^{3}\right)$ and $26.6 \%$. Compounds $1-5$ also showed antimicrobial activity against some fungi and gram-positive bacteria as shown in Table 1.

## Experimental

The IR and UV spectra were recorded on a JASCO A-102 and Shimadzu UV-220 spectrophotometer, respectively. ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectra were recorded on a JEOL GX-270 spectrometer. $J$-Values are given in Hz . FAB mass spectra were obtained on a JEOL HX-110 spectrometer using 2-nitrobenzyl alcohol as matrix. EI mass spectra (EIMS) were recorded on a JEOL DX303 spectrometer. Wako C-300 silica gel (Wako Pure Chemical) was used for glass column chromatography, and TLC was carried out on Merck silica gel GF254.

Isolation.-The sponge Niphates sp . ( 0.52 kg wet weight) collected by SCUBA off the Kerama Islands, Okinawa, was kept frozen until used. The methanol extract of the sponge was evaporated under reduced pressure and the residue ( 100 g ) was dissolved in a mixed solvent of ethyl acetate $\left(600 \mathrm{~cm}^{3}\right)$ and water $\left(600 \mathrm{~cm}^{3}\right.$ ) and was then partitioned between ethyl acetate ( 600 $\mathrm{cm}^{3} \times 3$ ) and water ( $600 \mathrm{~cm}^{3}$ ). The ethyl acetate-soluble material ( 8.43 g ) was partly ( 3.85 g ) subjected to silica gel flash column chromatography with gradient elution of methanol in chloroform ( $0-5 \%$ ) to give an active fraction $(1.21 \mathrm{~g})$, which was further purified by silica gel flash column chromatography and elution with gradually increasing amounts of ethyl acetate in hexane ( $0-100 \%$ ). The fraction ( 89.8 mg ) eluted with $20 \%$ ethyl acetate in hexane was further purified by HPLC [YMC-Pack AM-324 ODS, Yamamura Chemical; $10 \times 250 \mathrm{~mm}$; eluent,

Table 1 Antimicrobial activity of compounds 1-5

|  | MIC-values $\left(\mu \mathrm{g} / \mathrm{cm}^{3}\right)^{a}$ |  |  |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- |
|  | 1 | 2 | 3 | 4 | 5 |
| Candida albicans | 266 | 266 | $b$ | 33 | 266 |
| Cryptococcus neoformans | 133 | 133 | 133 | 16.5 | 133 |
| Paecilomyces variotii |  |  |  | 33 |  |
| Aspergillus niger |  |  |  | 133 | 133 |
| Staphylococcus aureus | 16.5 | 16.5 | 8.2 | 16.5 | 33 |
| Sarcina lutea | 133 | 133 | 133 | 33 | 66 |
| Bacillus subtilis | 266 | 266 | 266 | 133 | 133 |
| Escherichia coli |  |  |  |  |  |

${ }^{a}$ Minimum inhibitory concentration. ${ }^{b}$ No value here and elsewhere denotes 'inactive at $266 \mu \mathrm{~g} / \mathrm{cm}^{3}$ '.
acetonitrile-water-trifluoroacetic acid ( $50: 50: 0.1$ )] to yield niphatesines $\mathrm{E}\left(1,2.7 \mathrm{mg} ; t_{\mathrm{R}} 20.0 \mathrm{~min}\right), \mathrm{F}\left(2,5.4 \mathrm{mg} ; t_{\mathrm{R}} 32.8 \mathrm{~min}\right)$, $\mathrm{G}\left(3,5.4 \mathrm{mg} ; t_{\mathrm{R}} 44.0 \mathrm{~min}\right)$ and compound $5\left(8.5 \mathrm{mg} ; t_{\mathrm{R}} 28.0 \mathrm{~min}\right)$. The fraction ( 225.0 mg ) from the second silica gel column eluted with $50 \%$ ethyl acetate in hexane was separated by HPLC [YMC-Pack AM-324 ODS; $10 \times 250 \mathrm{~mm}$; eluent, acetonitrile-water-trifluoroacetic acid (30:70:0.1)] to give niphatesine $\mathrm{H}(4$, $9.5 \mathrm{mg} ; t_{\mathrm{R}} 15.6 \mathrm{~min}$ ) as well as the known compound niphatyne A $\left(7,16.5 \mathrm{mg} ; t_{\mathrm{R}} 35.6 \mathrm{~min}\right) .{ }^{6}$

Niphatesine $E$ 1.—An oil: $\lambda_{\max }(\mathrm{MeOH}) / \mathrm{nm} 262$ ( $\varepsilon$ 1900); $v_{\max }(\mathrm{KBr}) / \mathrm{cm}^{-1}$ 2910, 2850, 1680, 1200 and 1140; $\delta_{\mathrm{H}}\left(\mathrm{CDCl}_{3}\right)$ $8.84(1 \mathrm{H}, \mathrm{s}, 2-\mathrm{H}), 8.79(1 \mathrm{H}, \mathrm{d}, J 5.9,6-\mathrm{H}), 8.29(1 \mathrm{H}, \mathrm{d}, J 8.1,4-$ H), $7.83(1 \mathrm{H}, \mathrm{dd}, J 5.9$ and $8.1,5-\mathrm{H}), 7.36$ and $6.66(1 \mathrm{H}, \mathrm{t}, J 6.5$, $20-\mathrm{H}$; 1.4:1 ratio), 3.87 and 3.81 ( $3 \mathrm{H}, \mathrm{s}, \mathrm{OMe}$; 1:1.4 ratio), 2.99 $\left(2 \mathrm{H}, \mathrm{t}, J 6.6,7-\mathrm{H}_{2}\right), 2.58\left(2 \mathrm{H}, \mathrm{t}, J 6.6,8-\mathrm{H}_{2}\right), 2.06-2.18(4 \mathrm{H}, \mathrm{m}$, 11- and $19-\mathrm{H}_{2}$ ) and $1.20-1.43\left(14 \mathrm{H}, \mathrm{m}, 12-18 \mathrm{H}_{2}\right) ; \delta_{\mathrm{c}}\left(\mathrm{CDCl}_{3}\right)$ 152.1 and 151.1 (d, C-20; 1.4:1 ratio), 144.4 (d, C-4), 142.7 (s, C3), 140.7 (d, C-2), 140.5 (d, C-6), 125.9 (d, C-5), 83.9 (s, C-10), 76.0 (s, C-9), 61.6 and 61.2 (q, OMe; 1.4:1 ratio), 32.1 ( t$), 29.7$ (t), $29.3(\mathrm{t}), 29.2(\mathrm{t}), 29.0(\mathrm{t}), 26.8(\mathrm{t}), 26.2(\mathrm{t}), 25.6(\mathrm{t}), 20.0(\mathrm{t})$ and 18.6 (t); EIMS $m / z 314$ (5, M $^{+}$), 283 ( 80 ), 256 ( 10 ), 242 (35), 228 (11), 214 (12), 200 (16), 186 (20), 172 (18), 158 (37), 145 (30), 144 (25), 130 (13), 118 (12), 106 (28) and 93 (100) [HR-FAB-MS $m / z$, 315.2430. Calc. for $\left.\mathrm{C}_{20} \mathrm{H}_{31} \mathrm{~N}_{2} \mathrm{O}:(\mathrm{M}+\mathrm{H}), 315.2436\right]$.

Niphatesine F 2.-An oil; $\lambda_{\max }(\mathrm{MeOH}) / \mathrm{nm} 263$ ( $\varepsilon$ 3100); $v_{\max }(\mathrm{KBr}) / \mathrm{cm}^{-1}$ 2920, 2850, 1680, 1460 and $1200 ; \delta_{\mathrm{H}}\left(\mathrm{CDCl}_{3}\right)$ $8.73-8.75(2 \mathrm{H}, \mathrm{br}$ s, $2-\mathrm{and} 6-\mathrm{H}), 8.21(1 \mathrm{H}, \mathrm{d}, J 8.4,4-\mathrm{H}), 7.81-$ $7.86(1 \mathrm{H}, \mathrm{m}, 5-\mathrm{H}), 7.36$ and $6.66(1 \mathrm{H}, \mathrm{t}, J 6.2,22-\mathrm{H} ; 1.7: 1$ ratio $)$, 3.86 and $3.81\left(3 \mathrm{H}, \mathrm{s}, \mathrm{OMe}\right.$; 1:1.7 ratio), $2.94\left(2 \mathrm{H}, \mathrm{m}, 21-\mathrm{H}_{2}\right)$, $2.83\left(2 \mathrm{H}, \mathrm{t}, J 7.7,7-\mathrm{H}_{2}\right), 2.10-2.17\left(4 \mathrm{H}, \mathrm{m}, 14-\right.$ and $\left.17-\mathrm{H}_{2}\right), 1.67-$ $1.70\left(2 \mathrm{H}, \mathrm{m}, 8-\mathrm{H}_{2}\right)$ and $1.25-1.51(16 \mathrm{H}, \mathrm{br} \mathrm{s}, 9-13$ - and $18-20-$ $\mathrm{H}_{2}$ ); $\delta_{\mathrm{C}}\left(\mathrm{CDCl}_{3}\right) 151.9$ and 150.9 (d, C-22; 1.7:1 ratio), 144.7 (d, C-4), 142.9 (s, C-3), 141.7 (d, C-2), 139.5 (d, C-6), 126.3 (d, C-5), 80.3 and 79.9 (s, C-15 and -16), 61.5 and 61.2 ( $\mathrm{q}, \mathrm{OMe}$ 1.7:1 ratio), 32.9 ( t , $30.5(\mathrm{t}), 29.4(\mathrm{t}), 29.1(\mathrm{t}), 29.0(\mathrm{t}), 28.9(\mathrm{t}), 28.8(\mathrm{t})$, 28.7 (t), 28.3 (t) and 18.7 ( t ); EIMS $m / z 342$ (26, M ${ }^{+}$), 311 (28), 284 (17), 256 (10), 242 (34), 228 (21), 176 (28), 162 (20), 148 (26), 134 (28), 120 (32), 106 (17) and 93 (100) [HR-FAB-MS $m / z$, 343.2750. Calc. for $\mathrm{C}_{22} \mathrm{H}_{35} \mathrm{~N}_{2} \mathrm{O}:(\mathrm{M}+\mathrm{H})$, 343.2749].

Niphatesine G 3.-An oil; $\lambda_{\max }(\mathrm{MeOH}) / \mathrm{nm} 263$ ( $\varepsilon 1500$ ); $v_{\text {max }}{ }^{-}$ $(\mathrm{KBr}) / \mathrm{cm}^{-1} 2920,2850,1670,1200,1140$ and $720 ; \delta_{\mathrm{H}}\left(\mathrm{CDCl}_{3}\right)$ $8.70-8.76(2 \mathrm{H}, \mathrm{br} \mathrm{s}, 2-\mathrm{and} 6-\mathrm{H}), 8.17(1 \mathrm{H}, \mathrm{d}, J 8.1,4-\mathrm{H}), 7.77-$ $7.81(1 \mathrm{H}, \mathrm{m}, 5-\mathrm{H}), 7.20$ and $6.44(1 \mathrm{H}, \mathrm{d}, J 7.7,19-\mathrm{H} ; 3.2: 1$ ratio), 3.84 and $3.81(3 \mathrm{H}, \mathrm{s}, \mathrm{OMe} 3.2: 1$ ratio $), 2.82\left(2 \mathrm{H}, \mathrm{t}, J 7.7,7-\mathrm{H}_{2}\right)$, $2.33(1 \mathrm{H}, \mathrm{m}, 18-\mathrm{H}), 1.67-1.69\left(2 \mathrm{H}, \mathrm{m}, 8-\mathrm{H}_{2}\right), 1.25-1.32(18 \mathrm{H}, \mathrm{br}$ $\left.\mathrm{s}, 9-17-\mathrm{H}_{2}\right)$ and $1.05(3 \mathrm{H}, \mathrm{d}, J 6.6,18-\mathrm{Me})$; EIMS $m / z 318(6$, $\mathrm{M}^{+}$), 287 (100), 260 (111), 232 (38), 218 (7), 204 (10), 190 (9), 176 (8), 162 (9), 148 (11), 134 (8), 120 (18), 106 (45) and 93 (40)
[HR-FAB-MS $m / z, 319.2743$. Calc. for $\mathrm{C}_{20} \mathrm{H}_{35} \mathrm{~N}_{2} \mathrm{O}:(\mathrm{M}+\mathrm{H})$, 319.2749].

Niphatesine H 4.—An oil; $\lambda_{\max }(\mathrm{MeOH}) / \mathrm{nm} 262$ ( $\varepsilon 2500$ ); $v_{\text {max }}(\mathrm{KBr}) / \mathrm{cm}^{-1} 2920,2850,1200,1130$ and $720 ; \delta_{\mathrm{H}}\left(\mathrm{CDCl}_{3}\right) 8.78-$ $8.80(2 \mathrm{H}, \mathrm{br} \mathrm{s}, 2-\mathrm{and} 6-\mathrm{H}), 8.23(1 \mathrm{H}, \mathrm{d}, J 7.7,4-\mathrm{H}), 7.86-7.89$ ( 1 $\mathrm{H}, \mathrm{m}, 5-\mathrm{H}), 3.93(3 \mathrm{H}, \mathrm{s}, \mathrm{OMe}), 3.26\left(2 \mathrm{H}, \mathrm{br} \mathrm{s}, 20-\mathrm{H}_{2}\right), 2.98(2 \mathrm{H}$, $\left.\mathrm{t}, J 6.6,7-\mathrm{H}_{2}\right), 2.59(2 \mathrm{H}, \mathrm{t}, J 6.0), 2.08(2 \mathrm{H}, \mathrm{m}), 1.73(2 \mathrm{H}, \mathrm{br} \mathrm{s})$ and 1.16-1.29 (14 H, br s); $\delta_{\mathrm{C}}\left(\mathrm{CDCl}_{3}\right) 145.2(\mathrm{~d}, \mathrm{C}-4), 142.0(\mathrm{~s}$, $\mathrm{C}-3$ ), 140.8 (d, C-2), 140.5 (d, C-6), 126.4 (d, C-5), 84.2 ( $\mathrm{s}, \mathrm{C}-10$ ), 76.0 ( $\mathrm{s}, \mathrm{C}-9$ ), 61.4 ( $\mathrm{q}, \mathrm{OMe}$ ), 49.4 ( $\mathrm{t}, \mathrm{C}-20$ ), 31.9 ( t$), 29.0$ ( t ), 28.8 (t), 28.7 ( t$), 28.7$ ( t$), 28.7(\mathrm{t}), 28.4(\mathrm{t}), 27.8(\mathrm{t}), 26.1(\mathrm{t}), 23.6(\mathrm{t}), 19.8$ (t) and 18.4 (t); EIMS $m / z 316\left(5, \mathrm{M}^{+}\right)$, $285\left(95, \mathrm{M}^{+}-\mathrm{OCH}_{3}\right)$, 256 (18), 242 (12), 228 (15), 214 (20), 200 (21), 186 (22), 172 (18), 158 (45), 145 (30), 144 (20), 130 (10), 106 (25) and 93 (100) [HR-FAB-MS $m / z, 317.2600$. Calc. for $\mathrm{C}_{20} \mathrm{H}_{33} \mathrm{~N}_{2} \mathrm{O}:(\mathrm{M}+\mathrm{H})$, 317.2593].

Compound 5.-An oil; $\lambda_{\max }(\mathrm{MeOH}) / \mathrm{nm} 263$ ( $\varepsilon 3300$ ); $v_{\max }-$ $(\mathrm{KBr}) / \mathrm{cm}^{-1} 2920,2850,1680,1470$ and $1200 ; \delta_{\mathrm{H}}\left(\mathrm{CDCl}_{3}\right) 8.67-$ $8.86(2 \mathrm{H}, \mathrm{br} \mathrm{s}, 2-\mathrm{and} 6-\mathrm{H}), 8.21(1 \mathrm{H}, \mathrm{d}, J 6.2,4-\mathrm{H}), 7.82-7.84(1$ $\mathrm{H}, \mathrm{m}, 5-\mathrm{H}), 7.20$ and $6.46(1 \mathrm{H}, \mathrm{d}, J 8,18-\mathrm{H} ; 2.7: 1$ ratio $), 3.84$ and $3.80\left(3 \mathrm{H}, \mathrm{s}, \mathrm{OMe}\right.$; 1:2.7 ratio), $2.82\left(2 \mathrm{H}, \mathrm{t}, J 5.1,7-\mathrm{H}_{2}\right), 2.30-$ $2.35(1 \mathrm{H}, \mathrm{m}, 17-\mathrm{H}), 1.69(2 \mathrm{H}, \mathrm{m}, 8-2 \mathrm{H}), 1.25-1.32(16 \mathrm{H}, \mathrm{br} \mathrm{s}, 9-$ $16 \mathrm{H}_{2}$ ) and $1.04(3 \mathrm{H}, \mathrm{d}, J 6.6,17-\mathrm{Me}) ; \delta_{\mathrm{c}}\left(\mathrm{CDCl}_{3}\right) 157.1$ and 155.7 (d, C-18; 2.7: 1 ratio), 145.0 (d, C-4), 143.2 (s, C-3), 141.6 (d, C-2), 139.5 (d, C-6), 126.4 (d, C-5), 61.5 and 61.1 (q, OMe; 2.7:1 ratio), 34.8 ( t , 34.3 (d, C-17), 32.9 ( t$), 30.5(\mathrm{t}), 29.5(\mathrm{t}), 29.4$ ( t$)$, $29.3(\mathrm{t}), 29.1(\mathrm{t}), 29.0(\mathrm{t}), 27.2(\mathrm{t}), 27.0(\mathrm{t})$ and 18.2 and 17.6 (q, 17-Me; 2.7:1 ratio); EIMS $m / z 304$ ( $21, \mathrm{M}^{+}$), 273 (92), 246 (12), 232 (8), 218 (20), 204 (15), 190 (16), 176 (14), 162 (18), 148 (20), 134 (15), 120 (25), 106 (95) and 93 (100) [HR-FAB-MS $m / z$, 305.2584. Calc. for $\mathrm{C}_{19} \mathrm{H}_{33} \mathrm{~N}_{2} \mathrm{O}:(\mathrm{M}+\mathrm{H})$, 305.2593].

Hydrolysis of Compound 5.--Compound $5(1.5 \mathrm{mg})$ was treated with $3 \mathrm{~mol} \mathrm{dm}^{-3} \mathrm{HCl}-\mathrm{MeOH}\left(1: 1 ; 5 \mathrm{~cm}^{3}\right)$ under reflux for 20 h . After neutralization with aq. $3 \mathrm{~mol} \mathrm{dm}^{-3} \mathrm{KOH}$, the reaction mixture was extracted with dichloromethane $\left(5 \mathrm{~cm}^{3} \times\right.$ 3). The organic layer was partially evaporated, passed through an SEP-PAK cartridge $\mathrm{C}_{18}$ (Waters Associates), and eluted with acetonitrile-water ( $1: 1$ ) to afford the aldehyde $6(1.4 \mathrm{mg})$; $\delta_{\mathrm{H}} 9.61(\mathrm{~d}, J 1.9), 8.45(2 \mathrm{H}, \mathrm{br} \mathrm{s}, 2$ - and $6-\mathrm{H}), 7.52(1 \mathrm{H}, \mathrm{d}, J 7.9$, $4-\mathrm{H})$ and $7.21(1 \mathrm{H}, \mathrm{m}, 5-\mathrm{H})$; EIMS $m / z 275\left(11, \mathrm{M}^{+}\right), 246$ (29, $\mathrm{M}^{+}$- CHO), 218 (42), 204 (35), 190 (17), 176 (15), 162 (18), 148 (13), 134 (15), 120 (25), 106 (100) and 93 (85).

Reduction of Niphatesines $E$ (1) and $F$ (2).-To a solution of niphatesine $\mathrm{E}(1,2.0 \mathrm{mg})$ in methanol $\left(5 \mathrm{~cm}^{3}\right)$ was added sodium cyanoborohydride ( 1.5 mg ) and the pH -value of this solution was adjusted to 2 by addition of aq. $3 \mathrm{~mol} \mathrm{dm}^{-3} \mathrm{HCl}$. After the mixture had been stirred at room temperature for 6 h , the solvent was removed under reduced pressure and the residue was dissolved in ethyl acetate and washed with water. Evaporation of the organic layer afforded niphatesine $\mathbf{H}(4,2.0$ mg ), which was identified by comparison of TLC, ${ }^{1} \mathrm{H}$ NMR and EIMS data. Under the same conditions niphatesine F (2, 4.3 mg ) was treated with $\mathrm{NaBH}_{3} \mathrm{CN}(3.6 \mathrm{mg})$ to give niphatyne A (7, 4.1 mg ).

## Acknowledgements

We thank Mr. Z. Nagahama for his assistance in collecting the sponge. This study was supported in part by Grant-in-Aid (03151040) for Cancer Research from Ministry of Education, Science and Culture, Japan, and by Cooperative Research Program ('90-01) of the Research Center for Pathogenic Fungi and Microbial Toxicoses, Chiba University, Japan.

## References

1 J. Kobayashi, F. Itagaki, H. Shigemori, M. Ishibashi, K. Takahashi, M. Ogura, S. Nagasawa, T. Nakamura, H. Hirota, T. Ohta and S. Nozoe, J. Am. Chem. Soc., 1991, 113, 7812; H. Shigemori, S. Wakuri, K. Yazawa, T. Nakamura, T. Sasaki and J. Kobayashi, Tetrahedron, 1991, 47, 8429; J. Kobayashi, M. Tsuda, K. Agemi, H. Shigemori, M. Ishibashi, T. Sasaki and M. Mikami, Tetrahedron, 1991, 47, 6617; J. Kobayashi, M. Sato, M. Ishibashi, H. Shigemori, T. Nakamura and Y. Ohizumi, J. Chem. Soc., Perkin Trans. 1, 1991, 2609; J. Kobayashi, M. Sato, T. Murayama, M. Ishibashi, M. R. Wälchli, M. Kanai, J. Shoji and Y. Ohizumi, J. Chem. Soc., Chem. Commun., 1991, 1050; J. Kobayashi, H. Shigemori, M. Ishibashi, T. Yamasu, H. Hirota and T. Sasaki, J. Org. Chem., 1991, 56, 5221.

2 J. Kobayashi, T. Murayama, S. Kosuge, F. Kanda, M. Ishibashi, H.

Kobayashi, Y. Ohizumi, T. Ohta, S. Nozoe and T. Sasaki, J. Chem. Soc., Perkin Trans. 1, 1990, 3301.
3 A. R. Carroll and P. J. Scheuer, Tetrahedron, 1990, 46, 6637.
4 J. Kobayashi, T. Murayama, Y. Ohizumi, T. Sasaki, T. Ohta and S. Nozoe, Tetrahedron Lett., 1989, 30, 4833.
5 E. Pretsch, T. Clerc, J. Seibl and W. Simon, in Strukturaufklärung Organischer Verbindungen, Springer-Verlag, Berlin, 1981, pp. C195 and H175.
6 E. Quiñoà and P. Crews, Tetrahedron Lett., 1987, 28, 2467.

Paper 2/00888B
Received 19th February 1992
Accepted 20th February 1992

